FT-Raman Study of Hemoglobin with Quercetin and Naringin

S. Bakkialakshmi* & Jayoti Roy

Department of Physics, Annamalai University, Annamalainagar, Tamilnadu, India-608 002

Abstract:

In this article the FT-Raman spectra of human hemoglobin (HHb), Quercetin (Q) and Naringin (N) have been recorded separately. FT-Raman spectra for the complexes (1.) Hemoglobin-Quercetin and (2.) Hemoglobin-Naringin have also been recorded. The detailed interpretations of vibrational spectra of the complexes are furnished here, in terms of Potential energy distribution analysis.

Keywords: *Hemoglobin, Quercetin, Naringin, FT-Raman.*

I. INTRODUCTION

Vibrational spectroscopy is used extensively in organic chemistry, for the identification of functional groups of organic compounds, for studies in molecular conformation, reaction kinetic, etc. Flavonoids have emerged as potential therapeutic drugs and are effective against several diseases. Prominent among them, are free radical mediated [1, 2] and cell proliferative diseases [3, 4]. Flavones and flavonols are found in nature and have shown promising antioxidant and anticancer activity [5, 6]. Naringin has been reported to exhibit antioxidant, anti-inflammatory, anti mutagenic and anti carcinogenic effects and inhibits lipid per oxidation in biological membranes. It has antiviral and anti allergic actions and reduces the level of cytochrome P450 1A2 protein.

II. MATERIALS AND METHODS

FT-RAMAN spectra have been recorded on BRUKER RFS 27: Stand alone FT-Raman Spectrometer in the range 4000 - 100 cm⁻¹ at room temperature. The excitation line at 785nm has been taken from an Nd: YAG laser. Its scan number is 100, the resolution is 2cm⁻¹ and the sample is in solid phase.

III. RESULTS AND DISCUSSION

FT-Raman spectra of Hemoglobin have been depicted in Fig 1. FT-Raman spectra of Quercetin, Naringin and the complexes Hemoglobin - Quercetin and Hemoglobin - Naringin are given in Figs 2, 3, 4 & 5 respectively. FT-Raman absorption peak intensities of Hemoglobin before and after complex formation have been tabulated in Tables 1 & 2.

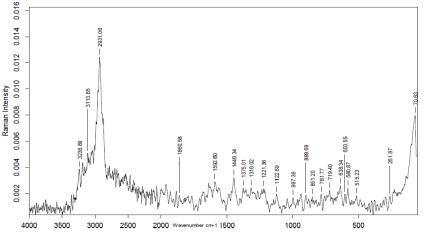
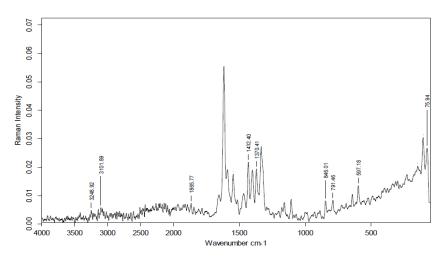
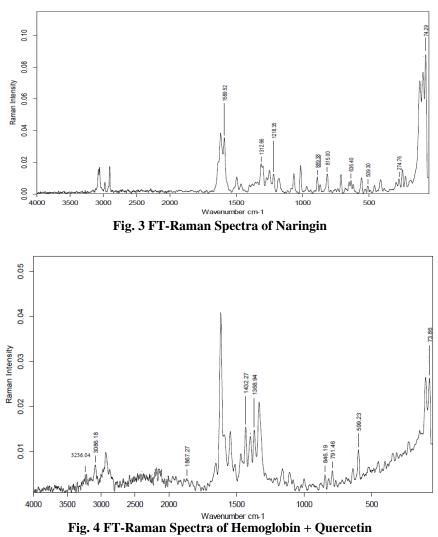
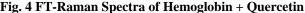


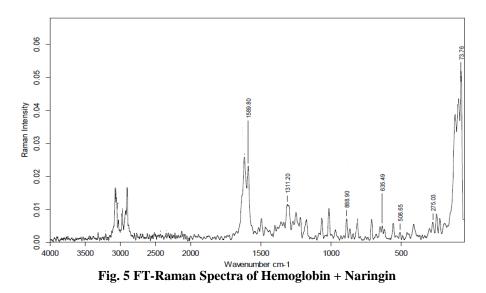
Fig. 1 FT-Raman Spectra of Hemoglobin











O-H vibration has been observed at 3236 ± 10 cm⁻¹. Aromatic ring vibrations were observed at 1432 ± 20 cm⁻¹. O-O vibrations have been observed at 846 ± 20 cm⁻¹. Aliphatic chain vibrations have been observed at 597 ± 20 cm⁻¹.

Intensities (cm ⁻¹)			Difference in	Tentative assignment
HbA	Q	HbA + Q	intensities prior to and after %	
3236.89	3248.92	3236.04	0.00094	O-H vibration
3113.65	3101.89	3086.18	0.00187	O-H vibration
1860.58	1865.77	1867.27	0.00212	C=C vibrations
1446.34	1432.40	1432.27	0.01217	Aromatic ring vibrations
1375.01	1370.41	1368.94	0.01233	Heme group vibrations, CH ₃ vibrations
851.20	846.01	846.19	0.00304	O-O vibrations
781.40	791.46	791.46	0.00375	C-C chain vibration
603.55	597.18	599.23	0.00845	aliphatic chain vibrations
70.63	75.94	73.86	0.0183	Lattice vibrations

Table 1 Difference In FT-Raman Absorption Peak Intensities of Hemoglobin Before and After Complex Formation with

Table 2 Difference in FT-Raman Absorption Peak Intensities of Hemoglobin Before and After Complex Formation with Naringin

Intensities (cm ⁻¹)			Difference in	Tentative assignment
HbA	Ν	HbA + N	intensities prior to and after %	
1592.60	1589.52	1589.8	0.02031	Aromatic vibration
1316.02	1312.66	1311.2	0.00917	aliphatic chain vibrations
1221.36	1218.35	1218.4	0.00525	C-C chain vibration
899.69	889.38	888.9	0.0054	O-O vibrations
638.34	636.40	635.49	0.00247	C-C chain vibration
515.23	509.3	508.65	0.00136	aliphatic chain vibrations
261.87	274.76	275.03	0.00438	aliphatic chain vibrations
70.63	74.29	73.76	0.04396	Lattice vibrations

IV. CONCLUSION

FT-Raman spectra of Hemoglobin, Quercetin, Naringin and the complexes (1.) Hemoglobin-Quercetin and (2.) Hemoglobin-Naringin were recorded and the detailed vibrational assignments were presented.

REFERENCES

- P.I. Oteiza, A.G. Erlejman, S.V. Verstraeten, C.L. Keen, C.G. Fraga, Flavonoidmembrane interactions: a protective role of flavonoids at the membrane surface. Clinical and Developmental Immunology 12 (2005) 19-25.
- [2] B. Sengupta, J. Swenson, Properties of normal and glycated human haemoglobin in presence and absence of antioxidant, Biochemical and Biophysical Research Communications 334 (2005) 954-959.
- [3] F.V. So, N. Guntherie, A.F. Chambers, M. Mousca, K.K. Carroll, Inhibition of human breast cancer cell proliferation and delay of mammary tumorigenesis by flavonoids and citrus juices, Nutrition and Cancer 26 (1996) 167-181.
- [4] G. Di Carlo, N. Mascalo, A.A. Izzo, F. Capasso, Flavonoids : old and new aspects of a class of natural therapeutic drugs, Life Sciences 65 (1999) 337-353.
- [5] B. Hendrich, R. Malon, A. Pola, Y. Shirataki, N. Motohashi, K. Michalak, Differential interaction of Sophora isoflavonoids with lipid bilayers, European Journal of Pharmaceutical Sciences 16 (2002) 201-208.
- [6] A. Arora, T.M. Byrem, M.G. Nair, G.M. Strasburg, Modulation of liposomal membrane fluidity by flavonoids and isoflavonoids, Archives of Biochemistry and Biophysics 373 (2000) 102-109.